Nature and Origin of Polymorphism in Feline MHC Class II DRA and DRB Genes¹

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Transcripts of the MHC class II DRA and DRB gene homologues of the domestic cat (Felis catus) were cloned and sequenced to compare the pattern and process of DR gene divergence. Homologous DRB exon 2 sequences from 36 feral domestic cats throughout the world plus from three species of Felidae (tiger cat, Iriomote cat, and Geoffroy's cat) were also determined. Limited variation in the domestic cat Feca-DRA gene was observed, but abundant variation in the Feca-DRB gene was seen comprising 61 distinct DRB alleles. Phylogenetic analyses resolved at least five monophyletic feline DRB allelic lineages (DRB*1 to *5), which are clearly distinct from those of human (HLA-DRB1 to 9 lineages), mouse (H-2FB b, u, f), and dog DRB alleles. Approximately 80% of individual cats contained three to six distinct DRB sequences, indicating that feline MHC maintains two to three DRB loci. Five cats had three DRB sequences in a single allelic lineage, indicating the occurrence of recent gene duplication of feline DRB genes. DRB sequences isolated from three exotic cats demonstrated close association with a particular domestic cat DRB lineage, suggesting that these allelic lineages are derived from common ancestral alleles that existed prior to the divergence of these feline species about 10 to 15 million years ago. Patterns of synonymous and nonsynonymous nucleotide substitution rates that occurred in Ag recognition sites (ARS) and nonrecognition (NAR) sites demonstrated a strong role of natural selection—positive selection for Ag recognition sites and negative selection for nonrecognition sites of feline DRB sequences—in the process of evolution of DR molecules. The Journal of Immunology, 1997, 158: 2822–2833.

he MHC encodes two categories of cell surface glycoproteins, which present short peptides to TCR (class I and class II molecules). Class I molecules, noncovalently associated with a β_2 -microglobulin, are expressed on the surface of all somatic cells. Class I molecules mediate the destruction of virus-infected cells and tumor cells by presentation of foreign-processed peptides to TCRs of CD8-positive CD4-negative cytotoxic T cells. Class II α - and β -chains are expressed on APC, such as B cells, dendritic cells, macrophages, and epithelial cells. These cell populations present peptides to TCRs of CD4-positive CD8-negative Th cells and activate an immune system through the secretions of specific cytokines (1).

The domestic cat is an important study model for neurologic disorders and for inherited diseases in humans (2). Domestic cats are also endemic with three pathogenic RNA virus infections—feline leukemia virus (3), feline immunodeficiency virus (4), feline infectious peritonitis virus (5). In addition, the cat family Felidae comprises 38 species and provides an important model for the pattern and processes of adaptive evolution (6, 7).

The cat MHC is located on the centromeric region of feline chromosome B2 (8, 9) and showed a moderate to high polymorphism in its class I and class II molecules by several parameters

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such as graft experiments (10), serologic analysis of cat pedigrees (10), RFLPs of feline class I genes (11), and sequence analysis of cDNA clones (12–14). A close examination of sequences of feline class I cDNA clones from three feline species (domestic cat, cheetah, and ocelot) representing the major taxonomic lineages of Felidae revealed two primary factors for generation and maintenance of polymorphic class I sequences, which include 1) positive selection pressure in favor of polymorphism or high heterozygosity in the Ag recognition site, and 2) periodic intragenic (interallelic) and intergenic (interlocus) segmental exchanges among class I genes (13, 14). The short polymorphic sequence motifs (20 to 23 bp) found in Ag-binding sites of class I molecules and other sequence motifs throughout class I sequences in the domestic cat were also found in the sequences of exotic cat species, suggesting a generation of these motifs in a common ancestor of these modern feline species and a persistence of the motifs for a long period of time (10) to 15 million years)—a transpecies mode of evolution (14).

We present here an analysis of class II DRA and DRB gene homologue variations in domestic cats, including 61 distinct DRB exon 2 sequences sampled from eight countries and four continents. The results reveal genetic monomorphism of DRA and extensive polymorphism and phylogenetic lineages of DRB allelic sequences reflecting ancient origin and sequence divergence of these molecules during the radiation of Felidae.

Materials and Methods

Animals

Samples from 37 feral domestic cat (Felis catus) were obtained from 11 locations in 8 countries, including Australia (AUS); Thailand (THAI); St. Petersburg, Russia (RUS STP); India (IND); Nicaragua (NCG); Central America (CAM); Buenos Aires, Argentina (ARG BA); Mendoza, Argentina (ARG MZ); Cordoba, Argentina (ARG CO); United Kingdom (UK); Maryland (USA MD); and Pennsylvania (USA PA) (see Table III) Genomic DNAs isolated from the tiger cat (Leopardus tigrina), Lti-6, and Geoffroy's cat (Oncifelis geoffroyi), Oge-1, and maintained in this laboratory were used for PCR analysis. PCR products of DRB exon 2-coding

¹ Tissue samples of endangered species were collected in full compliance with specific federal permits (CITES; Endangered and Threatened Species), issued by the U.S. Fish and Wildlife Service of the Department of Interior to the National Institutes of Health principal officer S. J. O'Brien.

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MhcFeca:

region from the Iriomote cat (Mayailurus iriomotensis) were a gift from F. Shinyashiki, (Okinawa International University, Ginowan, Okinawa).

Construction of cDNA library and isolation of feline DRA and DRB cDNA clones

mRNAs were extracted and purified from spleen cells of an adult domestic cat (FCA 021), cDNA was synthesized by a modification of the method by Gubler and Hoffman (15), using a cDNA synthesis kit (Pharmacia Biotech, Piscataway, NJ). EcoRI adaptor DNA (Stratagene, La Jolla, CA) was ligated with cDNA, and after phosphorylation of the 5' end by T4 DNA kinase, these cDNA were ligated with EcoRI sites of \(\lambda \) zap II vector (Stratagene) and in vitro packaged using GIGAPACK GOLD II extract (Stratagene). A total of 0.8 million original plaques were screened by 32P-labeled human HLA-DRA (16) and DRB1 (17) cDNA as probes.

PCR, cloning, and sequencing PCR-amplified DNA segments

PCR was performed to amplify exon 2 region (238 bp excluding primer sequences) from 100 ng genomic DNAs using oligonucleotides for human HLA-DRB genes: DRB219, 5'-CCACACAGCACGTTTCT/CTG-3'; and DRB61A, 5'-CCGCTGCACTGTGAAGCT-3' (18). PCR reactions were prepared in 50-µl volumes including 10 mM Tris-HCl (pH 8.3), 50 mM KCl, 1.5 mM MgCl₂, 200 µM each dNTP, 10 pmol of each primer, and 2.5 U of Taq DNA polymerase (Boehringer Mannheim, Indianapolis, IN). Thermal cycles were performed in a MJ Research PTC-100 thermocycler under the following conditions: 94°C for 3 min followed by 30 cycles of 94°C for 1 min, 57°C for 1 min, 72°C for 1 min, and 72°C for 10 min after the last cycle. Ten microliters of the PCR products were electrophoresed onto 3% agarose gel (2% NuSieve and 1% SeaKem GTG agarose, FMC Bioproducts, Rockland, ME) with a 123-bp DNA ladder marker (Life Technologies, Gaithersburg, MD) and visualized by staining with ethidium bromide. Samples containing DNA segments with the predicted size were treated with 1 U of Klenow fragment of DNA polymerase I at room temperature for 20 min to polish the ends. These samples were transferred into centricon 100 (Amicon Danvers, MA), and 1.5 ml of TE (10 mM Tris-HCl (pH 8.0) 1 mM EDTA) buffer were added and centrifuged at 6500 rpm for 10 min. Seven microliters of the sample were used to ligate with 100 ng of EcoRV-digested pBS SK+ vector (Stratagene) using 1 U T4 DNA ligase for 4 h at room temperature and transformed competent XL1-Blue Escherichia coli cells (Stratagene) by the following incubation: 4°C for 30 min, 42°C for 90 s, and 4°C for 2 min in Falcon 2059 tube (Becton Dickinson Labware, Lincoln Park, NJ). Bacteria and DNA solutions were then diluted 10 times by LB broth medium and incubated for 1 h at 37°C. Ten percent of these bacterial solutions were spread out onto LB agar plates containing ampicillin (100 μg/ml), X-gal (5-bromo-4-chloro-3-indolyl-β-p-galactoside), and IPTG (isopropylthio-β-D-galactoside); 50 μl of 20 mg/ml X-gal, and 10 µl of 10 mM IPTG were spread out per plate before plating transformed XL1 Blue cells. After incubating these plates at 37°C overnight, white-colored bacterial colonies were picked and the size of insert DNAs in the phagemid was monitored by PCR using a ScreenTest kit (Stratagene). E. coli cells harboring phagemids that contain the expected size of DNA insert were grown in 50 µg/ml ampicillin containing Super Broth (Quality Biological, Gaithersburg, MD) at 37°C overnight; phagemid DNA was isolated by Qiawell 8 System (Qiagen, Chatsworth, CA) according to the manufacturer's instructions and was sequenced by a method of cycle sequencing using M13 forward and reverse Dye Primer Prism Kit (Applied Biosystems, Foster City, CA) and an automated DNA sequencer (Applied Biosystems, model 373A). Each clone was sequenced in both orientations using M13 forward and reverse dye primers and confirmed both sequences were identical.

Sequence analysis

Sequence assembly and general sequence analysis were performed using the UWGCG program (19). Database search to identify similar sequences was performed by the NCBI BLAST program (20). Phylogenetic analyses were performed by the neighbor-joining method (21), and the protein sequence parsimony method (PROTPARS)3 (22) using PHYLIP program (22). Nonsynonymous and synonymous nucleotide substitution rates were calculated by Nei and Gojobori's method (23). All sequences studied here were submitted to GenBank by the following accession numbers.

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U51480; DRB*0102, U51482;
                                         DRB*0103.
                                                     U51483:
DRR*0101.
DRB*010401,
            U51484;
                     DRB*0105,
                                U51485;
                                         DRB*0106,
                                                    U51486:
DRB*0107.
           U51487:
                    DRB*0108.
                                U51488:
                                         DRB*0109.
                                                     U51489;
                                         DRB*0112.
                                                     U51492:
DRB*0110.
           U51490:
                    DRB*0111.
                                U51491:
                    DRB*0114,
DRB*0113,
           U51493;
                                U51494:
                                         DRB*0201.
                                                     U51496;
                                U51498:
                                         DRB*0204.
                                                     U51499:
DRR*0202
           1151497:
                    DRB*0203.
DRB*020501,
            U51500; DRB*0206, U51983;
                                         DRB*0207,
                                                    U51501:
DRB*0208,
           U51502;
                    DRB*0209,
                                U51503;
                                         DRB*0210,
                                                     U51504;
DRB*0211,
                                         DRB*0213,
                    DRB*0212,
                                U51506:
                                                    1151507:
           U51505:
DRB*0214, U51508 (exon 2 sequence of DRB*0214 cDNA clone);
DRB*0215.
           U51509:
                    DRB*0216.
                               U51510:
                                         DRB*0217,
                                                    U51511;
                                         DRB*0301.
DRB*0218,
           U51512;
                    DRB*0219,
                                U51513:
                                                     1151514
DRB*0302,
           U51515;
                    DRB*0303.
                                U51516;
                                         DRB*0304,
                                                     U51517:
                                         DRB*0307.
                                                     U51520:
DRR*0305
                    DRR*0306
                                1151519:
           U51518:
DRB*0308,
           U51521;
                    DRB*0309.
                                U51522;
                                         DRB*0310,
                                                     U51523:
DRB*0311.
           U51524:
                   DRB*0312,
                                U51525;
                                         DRB*0313,
                                                     U51526;
                                         DRB*0403.
                                                    U51529:
DRB*040101, U51527; DRB*0402, U51528;
DRB*0404.
           U51530; DRB*0405,
                                U51531;
                                         DRB*0501,
                                                     U51532:
DRR*0502.
           U51984: DRB*0503.
                               U51533: DRB*0504.
                                                    1351534
DRB*0505, U51535; DRB*0506, U51536 (exon 2 sequence of DRB*0506
cDNA clone); DRB*0507, U51537 (exon 2 sequence of DRB*0507 cDNA
clone); DRB*0508, U51538; DRB*0509, U51539; DRB*0510, U51540;
DRB*0511, U51541; DRB*0512, U51542.
MhcLeti:
DRB*0501, U51543; DRB*0502, U51544; DRB*0503, U51545.
MhcMair:
DRB*0401, U51546; DRB*0402, U51547.
MhcOnge:
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DRB*0201, U51548.

MhcFeca:

DRB*0214, U51574 (cDNA sequence); DRB*0506, U51575 (cDNA sequence); DRB*0507, U51573 (cDNA sequence).

MhcFeca:

DRA*0101, U51576 (cDNA sequence); DRA*0102, U51577 (cDNA sequence); DRA*0103, U51578 (cDNA sequence).

Results

Characterization of feline MHC DRA and DRB cDNA clones

Seven feline DRA and eight feline DRB cDNA clones were isolated by the secondary screenings of a spleen cell cDNA library from a domestic cat (Fca 021), using human DRA and DRB1 cDNA as probes. Three unique DRA and three unique DRB sequences were obtained from analysis of these cDNA clones. Derived cDNA sequences showed high identity to human DRA or DRB sequences, respectively, by database search. For instance, feline DRA clones showed 89.4 to 90.0% nucleotide sequence identity to human DRA and less than 70% identity to human DQA, DPA, and DNA, while feline DRB clones showed 84% sequence identity to human DRB, 69.9 to 73.5% to human DQB and DPB, 66% to human DOB (Tables I and II). Three unique sequences for both DRA and DRB cDNA clones were isolated from a single cDNA library (Figs. 1 and 2), suggesting that feline MHC has a minimum of two loci for both DRA and DRB genes. Alignment of nucleotide and deduced amino acid sequences with those of human DRA and DRB genes revealed that mature feline DRA and DRB molecules consist of 229 amino acids (aa) and 237 (aa), respectively. The feline DRA molecule has a 25 aa leader sequence, 84 aa α 1 and 107 aa α 2 domains, a 23 aa transmembrane, and a 15 aa cytoplasmic domain (Fig. 1). The feline DRB molecule has a 29 aa leader sequence, 95 aa \(\beta \)1 and 103 aa \(\beta \)2 domains, a 23 aa transmembrane domain and a 16 aa cytoplasmic domain (Fig. 2).

Eighteen polymorphic amino acid residues were found among these three domestic cat DRA molecules, and none of these residues was located in the α 1 domain, which is involved in the recognition of Ag and TCR (Fig. 3). In contrast, 33 of 35 polymorphic residues (94%) found in these three feline DRB sequences were located in the \beta1 domain, which is also involved in the recognition of Ag and TCR (Fig. 3). Nearly all of the polymorphic residues were located in its peptide-binding groove defined by an x-ray

³ Abbreviations used in this paper: PROTPARS, protein sequence parsimony method; ARS, antigen recognition sites; NAR, nonantigen recognition sites; dS, synonymous nucleotide substitution rates; dN, nonsynonymous nucleotide substitution rates; aa, amino acid; LB, Luria-Bertani.

Table I. The percentages of sequence identities among MHC class II α-chain coding sequences^a

	Feca-DRA*0101	Feca-DRA*0102	Feca-DRA*0103	Bola-DRA	HLA-DRA*0101	HLA-DQA1*0101	HLA-DPA1*0301	HLA-DNA
Feca-DRA *0101		98.5	96.7	88.2	89.7	63.3	68.4	65.9
Feca-DRA*0102	97.7		98.9	88.3	90.0	66.1	71.1	67.9
Feca-DRA*0103	94.5	98.6		87.5	89.4	63.9	69.3	66.9
Bola-DRA	84.9	84.5	83.7		85.7	64.3	68.9	67.0
HLA-DRA*0101	88.2	97.7	87 <i>.</i> 5	82.4		63.6	67.5	65.3
HLA-DQA1*0101	57.1	61.5	57.9	53.4	53.9		66.9	66.8
HLA-DPA1*0301	60.0	64.4	61.2	57.7	58.4	58.3		66.7
HLA-DNA	55.4	58.7	56.2	52.8	52.8	56.4	55.4	

⁴ Values in upper triangle and lower triangle represent percentages of nucleotide sequence identities, and amino acid sequence identities, respectively.

Feca-DRA*0101 Feca-DRA*0102 Feca-DRA*0103	-22 -1 TGCCGAAAACACCAAAGAAGAA.
Feca-DRA+0101 Feca-DRA+0102 Feca-DRA+0103	LEADER M A V S R V P V L G F C I M A L L M G P Q E S L A I K E E H 30 ATG GCC GTA AGT AGG GTC CCA GTG CTA GGA TTT TGC ATC ATG GCT TTA CTG ATG GGT CCT CAG GAA TCA TTG GCT ATC AAA GAA GAAG CAT 90 A. A. A. G TG
Feca-DRA*0101 Feca-DRA*0102 Feca-DRA*0103	V I I Q A E F Y L K P D S S G E F M F D F D G D E I F H V D 60 GTG ATC ATC CAG GCT GAG TTC TAT CTG AAG CCT GAC TCA TCA GGC GAG TTT ATG TTT GAC TTT GAT GGT GAT GAG ATT TTC CAC GTG GAT 180
Feca-DRA*0101 Feca-DRA*0102 Feca-DRA*0103	M E K K E T V W R L E E F G R F A S F E A Q G A L A N I A V 90 ATG GAA AAG GAG AGA GTG TGG CGG CTT GAA GAA TTT GGA CGT TTT GCC AGC TTT GAG GCC CAG GGT GCC TTG GCC AAT ATA GCT GTG 270
Feca-DRA+0101 Feca-DRA+0102 Feca-DRA+0103	ALPHA 2 D K A N L D I L I K R S N N T P N T N E P P E V T V L S N S 120 GAC AAA GCT AAC CTG GAC ATC TTG ATA AAG CGC TCC AAC AAC ACC CCA AAC ACC AAT GAA CCT CCT GAG GTG ACT GTG CTC TCA AAC AGC 360
Feca-DRA*0101 Feca-DRA*0102 Feca-DRA*0103	P V E L G E P N I L I C F I D K F S P P V F L P T W L R N G 150 CCT GTG GAA CTG GGA GAG CCC AAC ATC CTC ATC TGT TTC ATC GAC AAG TTC TCC CCA CCA GTG ATC AAT GTC ACG TGG CTT CGA AAT GGA 450
Feca-DRA+0101 Feca-DRA+0102 Feca-DRA+0103	K P V T T G V S E T V F L P R E D H L F R K F H Y L P F L P 180 AAG CCT GTC ACC ACA GGA GTG TCA GAG ACC GTC TTC CTG CCC CGG GAA GAC CAC CTT TTC CGC AAG TTC CAC TAT CTC CCT TTC CTA CCC 540
Feca-DRA+0101 Feca-DRA+0102 Feca-DRA+0103	S T E D V Y D C K V E H W G L D E P L L K H W E F D A P T P 210 TCG ACG GAG GAT GTC TAT GAC TGC AAG GTG GAG CAT TGG GGT TTG GAT GAG CCT CTC CTC AAG CAC TGG GAG TTT GAT GAC CCC 630
Feca-DRA*0101 Feca-DRA*0102 Feca-DRA*0103	TRANSMEMBRANE L P E T T E N V V C À L G L I V G L V G I I V G T I F I I K 240 CTC CCA GAG ACA ACA GAG AAT GTG GTG TGT GCC CTG GGC CTG ATC GTG GGT CTG GTG GGC ATC ATT GTC GGA ACC ATC TTC ATC ATC ATC ATC GCC CCA GAG ACC ACC TCC CACC GCC GCC GCC GCC GCC GCC GCC GCC
Feca-DRA*0101 Feca-DRA*0102 Feca-DRA*0103	G M R K V N A G E R Q G P L 254 GGC ATG CGC AAG GTC AAT GCT GGT GAA CGC CAA GGG CCT CTG TGA 765
Feca-DRA+0101 Feca-DRA+0102 Feca-DRA+0103 Feca-DRA+0101 Feca-DRA+0102	GGCATCCACAGGCAATGGCATTTACTAAAGAGAAGATCAATGAAGAAAATTTTGGCTTTAATAGCTTCAATGAGCCCAGGCAATACTCCAATTGTGCACCTCACAGAAGACAACTTTTCTCC C. G.A. GT. A. TG. TC. CT. T. A. T. T. T. C.C. C. G.A. GT. A. TG. TC. CT. T. A. T. T. AGCATTTTCAGCCCTTTAGCTGTTCTAAGAATAATGATGCCTGCTGATCTGTCCAGACTCTGACCTCATGTTATCCCCTGATTGTTGCCCCTTCTTGGGTATTGTTTCCTCCATTTTC 993
Feca-DRA*0103 Feca-DRA*0101 Feca-DRA*0102 Feca-DRA*0103	TATCTTCATTTTAATATCACCATGTAATGCCTCTGGAATAGACCCTAGAAGATCCTTTCTCTATTATGGAACCTTTTGAATGTTCAATATAGACATTTCTACATTTATTGTTTGGGT 1120

FIGURE 1. Nucleotide sequences of three MHC class II *DRA* cDNA clones of the domestic cat isolated from a single cDNA library. The complete nucleotide sequence and deduced amino acid sequence of *Feca-DRA*0101* allele is shown. Single letters and dots below the nucleotide sequence represent nucleotides that are, respectively, distinct from or identical to *DRA*0101*. Asterisks indicate missing sequences. Borders of each domain were assigned based upon sequence homology between *Feca-DRA*0101* and *HLA-DRA*0101*.

crystallographic model of HLA-DR1 MHC class II molecules (24). Nearly half of these sites (14 of 33) shared identical polymorphic residues found in human class II β -chain molecule (24).

PCR-amplified exon 2 region of the MHC class II DRB gene in natural populations of the domestic cats

To investigate these highly polymorphic regions, which consist of Ag-binding sites, we used PCR (25) to amplify the second exon,

which encodes the first extracellular domain of the *DRB* gene from 36 feral domestic cats in 11 geographic locales of eight countries (Table III). Sequence analyses of cloned *DRB* gene segments showed 61 distinct alleles by nucleotide sequences (Fig. 4); 59 alleles had a distinct amino acid sequence (Fig. 5).

In order to examine the relationship of these *DRB* sequences, we performed phylogenetic analysis for these feline *DRB* nucleotide sequences using the neighbor-joining method based on Kimura's

Table II. The percentages of sequence identities among MHC class II β-chain coding sequences^a

	Feca-DRB *0506	Feca-DRB *0507	Feca-DRB *0214	Dog MHC dw1	Bola-DRB3	HLA-DRB1 *0101	HLA-DQB1 *0501	HLA-DPB1 *01011	HLA-DOB
Feca-DRB*0506		95.5	94.7	86.2	84.9	84.0	73.5	70.4	65.9
Feca-DRB*0507	93.3		93.4	86.5	84.8	84.1	73.2	69.9	65.9
Feca-DRB*0214	92.2	89.1		86.4	84.4	84.1	73.5	71.0	65.7
Dog MHCdw1	81.0	83.6	83.2		84.3	86.2	<i>7</i> 3.8	72.6	66.4
Bola-DRB3	80.5	80.9	79.1	79.7		85.3	71.3	70.4	64.8
HLA-DRB1*0101	77.9	79.4	78.3	81.9	<i>77</i> .5		71.8	71.8	64.5
HLA-DOB1*0501	63.7	64.5	64.4	68.3	59.1	62.2		73.7	67.1
HLA-DPB1*01011	57.9	56.8	59.2	64.3	57.9	60.6	63.4		63.4
HLA-DOB	51.9	52.3	53.5	57.8	51.9	53.8	56.6	51.2	

^a Values in upper triangle and lower triangles represent percentages of nucleotide sequence identities, and amino acid sequence identities, respectively.

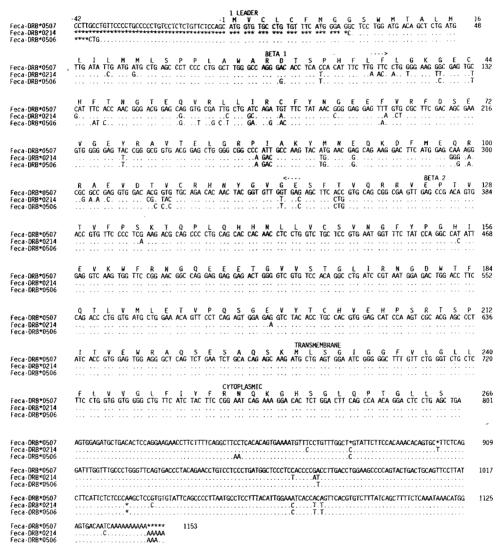


FIGURE 2. Nucleotide sequences of three MHC class II *DRB* cDNA clones of the domestic cat, isolated from a single cDNA library, FCA 021. Nucleotide and deduced amino acid sequences of the *Feca-DRB*0507* allele is shown. Single letters and dots below the sequence represent nucleotides that are, respectively, distinct from and identical to *DRB*0507*. Asterisks indicate missing sequences. Borders of each domain were assigned based on sequence alignment between *Feca-DRB*0507* and *HLA-DRB1*0101*. Arrows above the deduced amino acid sequence of β1 domain indicate PCR-amplified sequences excluding primer sequences.

two parameter distance (26). Six *DRB* sequences isolated from three nondomestic cat species—Geoffroy's cat (*O. geoffroyi*), tiger cat (*L. tigrina*), and Iriomote cat (*M. iriomotensis*)—were also dete

rmined and analyzed with representative sequences from human *HLA-DRB 1*, 3, 4, 5, 6, 7, and 9 lineages, mouse *H-2Eb* alleles, and dog-*DRB* sequences as outgroups.

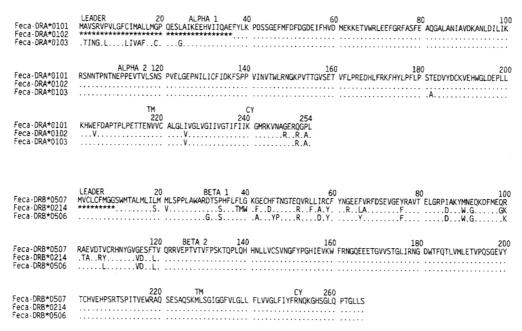


FIGURE 3. Aligned amino acid sequences of MHC class II *DRA* and *DRB* allelic sequences of the domestic cat, FCA 021. Amino acid sequences of *DRA*0101* and *DRB*0507* alleles are shown. Single letters and dots represent amino acid residues that are, respectively, distinct from and identical to the reference sequence. Asterisks indicate missing sequences.

The phylogenetic analysis (Fig. 6) revealed the following associations: 1) All feline sequences, both domestic and exotic species, are monophyletic (supported by a bootstrap score of 100%) and clearly divergent from human, mouse, and dog outgroup species. 2) The dog DRB sequences are the closest sequences to cat DRB (bootstrap for association equals 89%) as might be expected, since felids and canids are families of a single mammal order, Carnivora. 3) Five distinct clusters or allele lineages were resolved albeit with only modest bootstrap support and also confirmed by the PROTPARS method (data not shown); these groups are designated Feca DRB*1 to DRB*5. 4) Allele transcripts from DRB*1 to DRB*5 lineages are distributed throughout the world (Table III). 5) Limited DRB alleles derived from three nondomestic species are monophyletic for the species and align with one of the five Feca DRB lineages. In every case, the association derives from a deep ancestral node suggesting considerable mutation divergence of the DRB gene in domestic and nondomestic cats since divergence from their common ancestor. The species associations were: Geoffroy's cat (Onge-DRB*2), Iriomote cat (Mair-DRB*4), and tiger cat (Leti-DRB*5). These phylogenetic results suggest that the DRB allelic lineage was generated prior to the divergence of modern feline species from their ancestors, but subsequent to the split of Felidae from Canidae and noncarnivore species.

Based upon the phylogenetic assortment, we designate the sequences as MhcFeca-DRB*0101 to MhcFeca-DRB*0512 and used the simplified name, Feca-DRB*0101 to Feca-DRB*0512 in this paper, based on the proposal of nomenclature convention (27). The feline DRB sequences are likely alleles encoded by two or three DRB genes in domestic cats. Approximately 80% of domestic cats, in which more than four DRB clones were examined, contained three or more DRB allelic sequences and five cats had five or six DRB sequences (Table III). In five cases, we found that three sequences identified from a single cat fell in a same lineage. Of these, three cases showed three nearly identical sequences (0105/0106/0107 in FCA183, 0507/0508/0509 in FCA017, and 0208/

0209/0211 in FCA057), suggesting that each phylogenetic lineage is not restricted to a single locus, but recent gene duplication events subsequent to the generation of major phylogenetic lineages of feline DRB genes. The remaining two cases contained two closely related sequences plus one chimeric sequence. In one case (0104/0114/0115 in FCA177), 0114 sequence is a chimeric sequence between 0101 and 0104, indicating that two independent events-gene duplication plus interallelic (lineage) recombination—are possible mechanisms to explain these sequences. In the second case (0302/0305/0306), 0302 sequence is a chimera between 0303 and the last 30 bases of 0213, suggesting recombination between two lineages, in addition to a gene duplication event, could occur for the generation of this type of DRB polymorphism in feline MHC (Table III and Fig. 4). We also designated three DRB lineages from three exotic cat species as follows: MhcOnge-DRB*2 for Geoffroy's cat (O. geoffroyi), MhcMair-DRB*4 for Iriomote cat (M. iriomotensis), MhcLeti-DRB*5 for tiger cat (L. tigrina).

Positions of polymorphic sites in deduced amino acid sequences of feline DRB genes

In order to examine the nature of polymorphism found in the PCR-amplified exon 2 coding region of feline *DRB* genes, the positions of polymorphic amino acid residues in 61 cat *DRB* alleles were compared with those of 111 human *DRB* exon 2 sequences (provided by Dr. Colm O'hUigin, Max Planck Institute, Tübingen, Germany). Patterns of polymorphic sites were compared with the positions of Ag recognition sites (ARS), designated by ARS definition based on x-ray crystallography of *HLA-DR1* class II molecules (24) (Fig. 7). For 111 human *HLA-DRB* alleles, 124 polymorphic amino acid residues were found in a 95 aa segment (95 sites) of the first extracellular domain, (1.30 polymorphic residues/site) and of these 63 polymorphic residues are located on defined (22 sites) ARS, respectively (Fig. 7). These results indicated significant accumulation of polymorphic sites in ARS of human *HLA-DRB* molecules (2.86 polymorphic residues/site in

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Table III. DRB alleles of domestic cats^a

		No. of DRB	No. of DRB			DRB Alleles Present		
Cat Code	Location	Clones Sequenced	Alleles Found	DRB*1	DRB*2	DRB*3	DRB*4	DRB*5
FCA168	AUS	1	1			0304		
FCA169	AUS	4	3		0201, 0204	0308		
FCA170	AUS	1	1	0107				
FCA171	AUS	3	2	0102				0511
FCA172	AUS	3	2			0301, 0304		
FCA174	AUS	2	1	0111				
FCA175	AUS	3	2	0108	0206			
FCA177	AUS	4	3	0103, 0113, 0114				
FCA178	THAL	3	3	0104		0310	0403	
FCA179	THAI	4	3		0213			0501, 0503
FCA180	THAL	1	1					0502
FCA181	THAI	4	3	0107	0202	0304		
FCA182	THAI	5	5		0210, 0216	0304, 0307		0501
FCA183	THAI	8	4	0104, 0105, 0106	0207	,		
FCA184	THAI	2	2			0304, 0307		
FCA144	RUS STP	4	2	0107	0201			
FCA145	RUS STP	1	1		0212			
FCA120	IND	3	2	0107		0304		
FCA150	CAM	5	5	0103, 0107	0215		040101, 0402	
FCA151	NCG	5	4		0201	0302, 0305, 0306		
FCA153	ARG BA	3	2	0102, 0107		0304		
FCA154	ARG BA	3	3	0107				0504, 0512
FCA155	ARG MZ	2	2	0101, 0107				,
FCA156	ARG CO	4	3	0109, 0110				0505
FCAOT3	UK	3	3	0103, 0107			0405	
FCATA2	UK	2	2			0303, 0304		
FCACW2	UK	2	2	0112	0219	,		
FCA011	USA MD	4	2		0203		0403	
FCA017	USA MD	7	5	0107		0312		050701, 0508, 0509
FCA021	USA MD	8	3		0214			0506, 050702
FCA039	USA MD	4	2		0207	0311		
FCA042	USA MD	8	1				040101	
FCA057	USA MD	11	6		0208, 0209, 0211	0311	040102, 0404	
FCA062	USA MD	4	4		020501, 0209	0303, 0304	3.0.02,0104	
FCAC114		3	2		020502, 0218			
FCAPA107		8	5		0209	0310, 0311		050702, 0510
FCAPA116		2	2			0310, 0313		5557 OZ, 00.0

^a Geographic locations for sample collections were indicated as abbreviated letters as follows: ARG BA, Argentina Buenos Aires; ARG CO, Argentina Cordoba; ARG MZ, Argentina Mendoza; AUS, Australia; CAM, central America (exact location was not known); IND, India; NCG, Nicaragua; RUS STP, Russia, St. Petersburg; THAI, Thailand, UK, United Kingdom; USA MD, United States of America, Maryland; USA PA, United States of America, Pennsylvania.

defined ARS). Sixty-three feline *DRB* alleles display 115 polymorphic residues in a 78 aa segment of first extracellular domain-coding region (1.47 polymorphic residues/site). Of these, 58 were in the 22 positions of the defined ARS (2.6 residues/site for the defined ARS). These results indicated that distributions of polymorphic residues in cat *DRB* is rather similar to human *DR* molecules, except that Feca *DRB* alleles have more polymorphic residues than human *DRB* molecules at the last three ARS (positions 78, 81, and 86).

Nucleotide substitution patterns of ARS and non-ARS of feline DRB molecules

The pattern of nucleotide substitution in feline *DRB* sequence at ARS and non-ARS sites defined by the x-ray crystallographic model (28–30) were examined. We calculated synonymous (dS) and nonsynonymous (dN) nucleotide substitution rates for the ARS and non-ARS in all pairwise comparisons of 49 *DRB* sequences (13, 14, 30) using Jukes and Cantor's genetic distance (31). Average dS and dN estimates for each feline *DRB* sequence comparison are presented in Figure 8. With a few exceptions, dN values were greater than dS values for ARS,

while the reverse is true for non-ARS sites (Fig. 8). These results are parallel to the analyses of MHC class I genes (13, 14, 30), although the ratios of dN and dS values in the ARS of class I molecules are much higher than those of class II DRB molecules (dN = 15.2, dS = 7.2 for domestic cat class I molecules' ARS (13). This was caused by lower average dN value in ARS sites of feline DRB molecules compared with dN values in ARS sites of human DRB and mouse Eb molecules (18.5 for feline DRB, and 45.7 and 41.5 for human DRB and mouse Eb molecules, respectively) since the dS value in the ARS of feline DRB molecules is equivalent to values of two other species (13.3 for feline DRB, 15.0 for human DRB, and 11.6 for mouse Eb molecules) (32 and this study).

Discussion

Sequence analyses of domestic cat MHC class II *DRA* and *DRB* genes showed remarkable similarities to the human HLA class II *DR* system in the quantity and quality of polymorphism for *DRA* and *DRB* class II genes. In human HLA, class II *DRB* genes are highly polymorphic with some 126 *DRB* alleles identified to date (Table IV). In contrast,

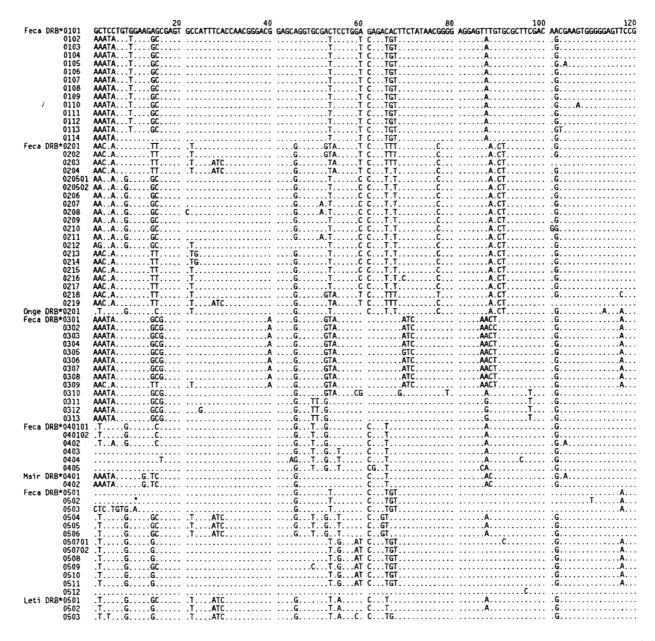


FIGURE 4. A complete list of nucleotide sequences of β1 domain coding region of feline MHC class II *DRB* genes. Single letters and dots represent nucleotides that are, respectively, distinct from and identical to *DRB*0101*. The allelic numbers for domestic cat and exotic cats were assigned based upon five major lineages of *DRB* alleles found by phylogenetic analysis (Fig. 6) and sequence alignments of amino acid sequences by PILEUP program of UWGCG (Fig. 5). MHC abbreviations used for species were *Feca, Felis catus* (domestic cat), *Onge, Oncifelis geoffroyi* (Geoffroy's cat), *Mair, Mayailurus iriomotensis* (Iriomote cat), *Leti, Leopardus tigrina* (tiger cat).

the *DRA* gene, which encodes the other half of a heterodimeric *DR* class II molecule, has only two alleles in human populations (33). The domestic cat MHC encodes at least 61 *DRB* alleles in worldwide populations of the domestic cat, while we resolved only one *DRA* allele by the sequences in exon 2, although nucleotide substitutions were found in other regions (Fig. 1). Similarly, a majority of mammalian MHC studied so far maintained a number of *DRB* alleles (Table IV), while *DRA* gene is monomorphic (34).

A comparative analysis of the positions of polymorphic residues in the first extracellular domain of the *DRB* molecule in humans and domestic cats showed the extraordinary similarity

of positions of polymorphic residues facing defined ARS (Fig. 7). Furthermore, examination of the patterns of nucleotide substitutions in ARS, and other regions in exon 2 of these feline DRB genes indicates selection pressures observed in feline class II genes, which include positive selections for amino acid substitutions in the Ag-binding site, but negative selection pressure against codon changes in the framework and NAR of DRB molecules. This evidence seems to suggest strong functional constraints for the patterns of polymorphism in Ag-binding sites of DRA and DRB molecules during long divergence time between humans and the domestic cat.

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	140	160	180	200	220	238
Feca DRB*0101 0102	GGCGGTGACGGAGCTGGGGC	GGCCCACTGCCAAGTACTGG	AACGAGCAGAAGGACCACC!	GGAGCAGGAGCGGACAGCGG	16GACCGGATCTGCAGACAC	AAL I ALGGI GI I GGI GAG
0103						
0104		AGAC				
0105	T	AGAC				• • • • • • • • • • • • • • • • • • • •
0106 0107		AGAU				
0108				AC		,
0109						
0110 0111						
0112				******************		
0113						
0114 Feca DRB*0201	AAT.A	TGACT.		GG.ACG.C.A		
0202	AAT.A	, AGAC		ACG.C.A	AC.GT	
0203	AAT.A	AGAC	T.GTT.A.	G.AACG.C.A	AC.G	
0204		AGAC			ACCG	
020501 020502	•••••	TGACC	T.GT.A.	AC	T C	
0206		TGACC	T.GATT.A.	CAC	TC	
0207		TGACC	T.GTT.A.	AC	TT	
0208 0209		TGACC		AC		
0210			T.G	AC	T T	
0211		TGAC.GC	T.GTT.A.	AC	TT	
0212			g			
0213 0214			GTT.A.	GG.A	TA	
0215			GTT.A.			
0216		AGAC	GTT.A.	GG . A	, TA ,	
0217 0218	• • • • • • • • • • • • • • • • • • • •	AGAC C	GTT.A.	GG.A	<u>.</u> <u>T</u> A	
0218	A AT A	AGAC	T.G. T. TT A	G. A	T T	
Onge DRB*0201		T	GTT.,A.	G	T,TAT	
Feca DRB*0301 0302		.AAGAC	GTTA.	GA	<u>T</u> A	····· <u>ī</u> ····· <u>ū</u> ····č
0303		TAGAC		GA	TA	ππс
0304		AGAC	GTTA.	GAC	T T	
0305			GTTA.			
0306 0307			G T T A	GAC	TT	•••••
0308		AGAC	GTTA.	GAC	TT	
0309		AGAC		GAC		
0310 0311	A	TGAC	T.GTGT.A.	Δ 1 Ω1 Δ	TC	TTC
0312		TGAC	GTTT.A.	A CG. C. A.	AC. G.	TTC
0313	<u></u>	TGAC	GTTT.A.	A,CC.A	AC.G	TTC
Feca DRB*040101 040102		TCT.	GTTA.	GTCA		
0402		T	GTTA.	GTC		
0403	<u>I</u>			GTC	ACCTA	<u>T</u> c
0404 0405	······	AGAC	GIIA.	GTC	ACCTA	TTC
Mair DRB*0401		TTG	GTTA.	GTC	AC.TA	TTTC
0402		TTG	GTTA.	GTC	AC.TA	
Feca DRB*0501 0502	· · · · · · · · · · · · · · · · · · ·	AGAC	GTT.A.	ACG.C.A	AC.G	
0502		AGAC	GTT.A.		AC G	
0504	AT	***************************************	GTT.A.	TA.ACG.C.A	AC.G	
0505 0506	T	ACAC	G	TA.ACG.C.A	AC.G	
050701		AGACAT.	υ	ACG.C.A AAGCG.C.A	ACCC	TC
050702		TAT.		AAGCG.C.A	AC.G.G	
0508			TT.A.	AAGCC.A	AC.G.G	***************************************
0509 0510	.c	GTAT.		AAGCG.C.A	AC.G.G	• • • • • • • • • • • • • • • • • • • •
0511		AGACAT.	TT.A.		AC.G.G.	T . C
0512		TCT.	TT.A.	ACC.A	AC.G	• • • • • • • • • • • • • • • • • • • •
Leti DRB*0501 0502	e	TGAC		G.ACG.C.A	AC.G	• • • • • • • • • • • • • • • • • • • •
0502		T	GTTT.A.	ACG.C.A	AC.G	T

FIGURE 4. (continued)

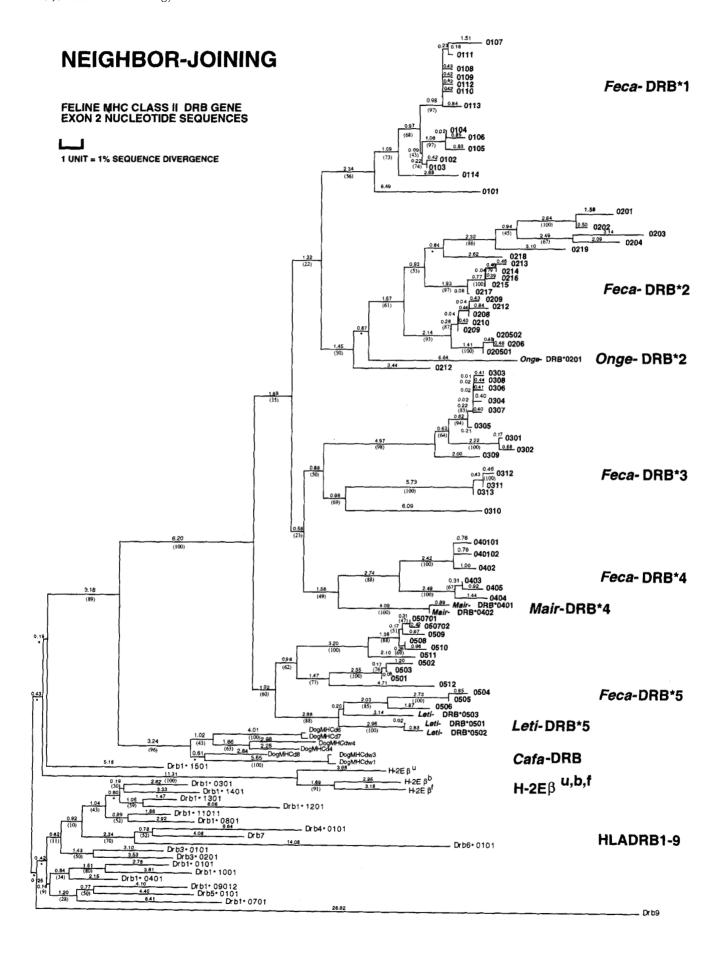
The occurrence of three distinct cDNA clones for both DRA and DRB genes from a single spleen cell cDNA library plus the occurrence of five or six DRB sequences in five cats (Table III) suggests that feline MHC contains at least two DRA loci and three DRB loci. Phylogenetic analysis for feline DRB sequences revealed five lineages of DRB allelic sequences that originated after the divergence of Felidae from carnivore families (e.g., canids), but before the more recent emergence of three major taxonomic groups of Felidae—ocelot, domestic cat, and pantherine lineages (6, 7). Identification of three sequences within a single phylogenetic lineage identified from a single cat in five cases suggests two possible events, 1) recent gene duplication, and 2) segmental exchanges between two lineages. Of these two possible events, recent gene duplication events of DRB genes is the most likely explanation, since apparent evidences of segmental exchanges between two lineages were rare. Only Feca DRB*0302 in these five cases has chimeric structure between Feca*0303 and the last 30 bases of 0213. The close association of *DRB* alleles of three nondomestic species (Geoffroy's cat, Iriomote cat, and tiger cat) with domestic cat *DRB* lineages suggests persistence of MHC allelic lineages through multiple speciation events (a trans-species mode of evolution) during the Felidae radiation since the last 10 to 15 million years.

Apparent mosaic sequences observed in feline MHC class I genes, which suggested inter- and intralocus segmental exchanges as a primary factor for the evolution of class I genes, were rarely seen in feline *DRB* exon 2 sequences, suggesting different modes of evolution operate diversification of feline MHC class I and class II *DRB* genes. Nonetheless, the transspecies pattern of allele divergence, the abundant allelic diversity, and the positive selection in favor of divergence observed in the ARS are consistent with an important role for the *DR* loci in immune response to pathogenic agents in ancient and modern species of felids.

Feca DRB*0101	20	40 RHFYNGEEFVRFDNEVGEFR	60	79
0102	NWI V E A	.CYS	AVIELGRIARIWNEURDHL	EVERTAVORICKHNYGVGE
0103	NMI A F V	.CYS		A
0104	NMI A F.V	. c YS	D 6	
0105	NML.AF.V	.cYS	D G	
0106	NML.AF.V	.CYS	P D 6	
0107	NML.AF.V	.CYS		
0108		.CYS		
0109	NML.AF.V	.CYS		
0110	NML.AF.V	.CYS.M		
0111	NML.A	.CYS		N
0112	NML.AF.V	.CYS	Q	
0113	NML.A	.cys		
0114	NMF.V	.cYS	Y.	WF
Feca DRB*0201	TMF	.FRLAS	s n i fw	CV AF TV
0202	TM E D V V	.FRLAS		V AE TUE
0203	TM F VD D V V	.FRLA	S D C EM	DVUAE TV EN
0204	TM F VD P V V	.YRLAS	D G EV	DVHAF TV EN
0205	IMG.AR. F.A	.YRLAS	D H G FM	T WI FO
0206	IMG.AR.F.A	.YRLAS	D. H. G. FM	D.T. WI FD
0207	IMG.AR.OF.A	.YRLAS	DHGFM	TWF
0208	IMG.A.SR.OF.A	.YRLAS	DHGFM	TWF
0209	IMG.ARF.A	.YRLAS	DHGFM	TWF
0210	IMG.A	.YRLAG	DHGFM	TWF
0211	IMG.AR.QF.A	.YRLAS	DG.HGFM	TWFS.
0212	VMG.A	.YRLAS		ASSFD
0213	TMFDRF.A	.YRLAS	D.RGFM	.GKVD
0214	TMFDR.,F.A	.YRLAS		.GKVD
0215	TMFRF.A	.YRLAS	DGFM	.GKYVD
0216	TMFRF.A	.YSRLAS	DGFM	.GKYVD
0217	TMFRF.A	.YRLAS	DGFM	.GKY
0218	IMFRY.V	.FWLASS.	DHGFM	TWE
0219	IMFYPRY.V	.FRLA	SDGLFM	.RKWF
Onge DRB*0201	F.G.1	.YRLASKY.	IL.GLYM	GWYY
Feca DRB*0301	NM A D V	YHNLSY.	D GL VM	EK A EU
0302		YHNLSY.		
0303		YHNLSY.		
0304	NMARY	YHNLSY.	DGI. YM	.FTWF
0305	NM . A R . Y	CHNLSY.	D. GL YM	FT WF
0306		YHNLSY.		
0307		YHNLSY.		
0308		YHNL\$Y.		
0309	TMFRY	YHNLSY.	DGLYM	.ETWF
0310	NMARY.R	VYS		.RRFD
0311	NMAR.W		DGLFM	K.AETVFD
0312	NMARR.W		DGLFM	K.AETVFD
0313	NMAR.W		DGLFM	KETVFD
C DDD+0401	F 0 F D D	v v 5	T 1 01 VM	EC TV
Feca DRB*0401 0402		.YYS		
		.YYS		
0403 0404	U	.YYL.S	D CI VM	ES TV V
0405		GYHS	n cl VM	ES TV VO
Mair DRB*0401			VF.EGLYM	
0402	NM.R	Y Y S	V FF GI YM	FS TY FD
0 102				
Feca DRB*0501		.CY.	DGFM	K.AETV
0502		.CW.Y.	MDGFM	K.AETV
0503	SCG	.CY.	DGFM	K.AETV
0504	F.G.AYPRD	.YYS		D.K.AETVD.
0505	F.G.AYPRD	.YYS		D.K.AETV
0506	F.G.AYPRD	,YYS	DGFM	K.AETLVD
0507	F.G.G	.c	FM	R.AETV
0508	F.G.GI	.ç	<u>I</u> <u>M</u> <u>EM</u>	RETV
0509	F.G.A	.çy.	RIMEM	K.AEIV
0510	F.G.G	.cy.	P	KEIV
0511	F.G.GI	.cs	pm <u>FM</u>	K.ALIVVD
0512	T.O.A. VD		řřr.žř.hw	KEIV
Leti DRB*0501	r.u.A17KD	.yyş		DV AC TV
0502	1.41.41171717	.YYS	ייםעה.בולי אם היים ל	N AE TV V
0503	F.u.u17K			V.

FIGURE 5. Feline MHC class II *DRB* allele. Amino acid sequences of PCR amplified first extracellular domain coding region of *DRB* genes. The position 1 of this figure corresponds to position 9 of β 1 domain (See Fig. 7).

FIGURE 6. Neighbor-joining tree using 238-bp nucleotide sequences of β1 domain coding region in MHC class II *DRB* genes based on Kimura's two-parameter distance. Phenetic distance branch length (percent sequence divergence) is indicated above each branch, numbers in parentheses are bootstrap percentages (out of 100 iterations) in support of each node. MHC abbreviations used for species are indicated as follows: *Feca, Felis catus* (domestic cat), *Onge, Oncifelis geoffroyi* (Geoffroy's cat), *Mair, Mayailurus iriomotensis* (Iriomote cat), *Leti, Leopardus tigrina* (tiger cat), *Cafa, Canis familiaris* (domestic dog). *H-2E* β and *HLADRB* are used for murine and human class II β-chain sequences, respectively.



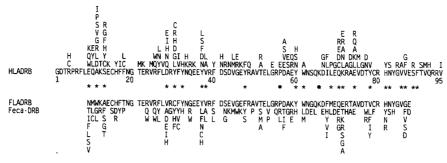


FIGURE 7. Distributions of highly polymorphic and conserved amino acid residues in β 1 domain of feline and human *DRB* class II molecules. Consensus amino acid sequences in first extracellular domain of *DRB* class II molecule for human (*HLADRB*) and domestic cat (*FLADRB*) were shown. Each consensus amino acid sequence for *HLADRB* and *FLADRB* does not represent an individual molecule. ARS based on x-ray crystallographic analysis of human class II *HLA-DR1* molecule (24) are indicated by asterisks. Single letters above human and below cat *DRB* sequences represent polymorphic amino acid residues.

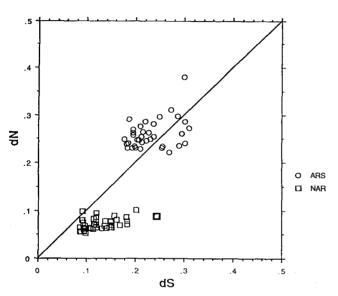


FIGURE 8. Synonymous (noncodon altering) and nonsynonymous (codon altering) nucleotide substitution rates in the ARS, NAR defined by an x-ray crystallographic structure of *HLA-DR1* class II Ags. Fortynine *DRB* sequences were analyzed by pairwise comparisons using the method of Nei and Gojobori (23) to calculate dS and dN for their ARS (circles) and NAR (rectangles). Each symbol represents the average of all pairwise comparisons that measure the number of nonsynonymous or synonymous substitution/total number of sites for potential nonsynonymous or synonymous substitution for each sequence.

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Table IV. Number of DRB allelic sequences in mammals

Spec	cies				
Common name	Species name	No. of Individuals Studied	No. of DRB Alleles	Reference No.	
Human	Homo sapiens	251	126	33	
Common chimpanzee	Pan troglodytes	64	85	35	
Gorilla	Gorilla gorilla	19	40	35	
Domestic cat	Felis catus	37	61	This study	
Cattle	Bos primigenius taurus, Bos primigenius indicus	Not known	64	36–43	
Red deer	Cervus elaphus	50	34	44	
Pig	Sus scrofa	12	9	45, 46	
Sheep	Ovis aries	15	13	47	
Goat	Capra hircus	25	21	47	
European moose	Alces alces	30	10	48	
North American moose	Alces alces	19	10	48	

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